Adulticide Efficacy of Artemisia vulgaris L. against Aedes aegypti L.

Potensi Adultisida dari Artemia vulgaris L. terhadap Aedesagypti L.

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ABSTRACT

Aedes aegypti is the vector of various arthropod-borne diseases such as dengue fever, chikungunya and currently, zika. This study aimed to evaluate Artemisia vulgarisas other adulticides for controlling Ae. aegypti. Indonesian ministry of health hasbeen reported that more than 70.000 dengue fever human cases have occurred in 2015 covering 34 provinces which mean that dengue fever has spread now into all national territories. Eradication of Aedes still largely depends on insecticides, which is the most cost-effective strategy, and often inefficient due to resistance development in exposed Aedes population This study was designed to use of Centers for Disease Control and Prevention (CDC) bioassay standard. CDC bottles were coated with the ethanolic solution of extract with a concentration of 10, 50, 100, 500, 1000, 5000, 10000, 50000, and 100000 µg per bottle. F0 mosquitoes were used for all experiments. Death and surviving mosquitoes were evaluated based on CDC standard assay. The test was performed with 10-25 adult mosquitoes every bottle and each concentration was repeated in triplicates. The results showed that LC_{50} and LC_{90} were 5790 µg and 52110 µg respectively after 120 minutes exposure to the extract. There was no mortality in ethanol control group. A. vulgaris significantly (P < 0.05) have adulticidal activity against Ae, aegypti. However, A. vulgaris have been reported to have larvacidal activity against Ae. Aegypti. These results indicated clearly that A. Vulgaris might act as the candidate of bioinsecticides for controlling Ae. aegypti.

Keywords: Aedes aegypti, Artemisia vulgaris, adulticide, bio insecticide.

INTRODUCTION

Mosquitoes are the vector of various disease such as malaria, chikungunya, dengue fever, vellow fever, Japanese encephalitis, filariasis, schistosomiasis and currently zika (Govindarajan et al., 2011). There are 111 genera and 137 subgenera containing 3,517 species of mosquitoes in worldwide (Becker et al., 2010). Aedes aegypti is a major vector of arthropod-borne disease, primary vector responsible for dengue fever, yellow fever, chikungunya, and Zika fever (Ajaegbu et al., 2016). Dengue is major public health problem globally (Samuel & Tyagi, 2006) over 3.97 billion people in 128 countries at risk of disease. At 2010 approximately there were 390 million cases (Sayono et al., 2016). Indonesian ministry of health has been reported that more than 70.000 dengue fever human cases have occurred in 2015 covering 34 provinces which mean that dengue fever has spread now into all national territories(Hamid et al., 2017). While vaccine of dengue fever is still developing with active research, the only way to prevent and

control dengue virus transmission by controlling the vector (Wuliandari *et al.*, 2015, Elumalai *et al.*, 2016). The application of chemical and synthetic insecticide has been long utilized, there were several problems such as insecticidal resistance, environmental pollution, and harmful impact on human and other organisms (Cheah *et al.*, 2013).

Mosquitoes have been resistance to synthetic and chemical insecticide in several places in Indonesia, as in Central Java province (Sayono et al., 2016), Yogyakarta province (Wuliandari et al., 2015), Bali (Hamid et al., 2017). Bandung, Surabaya, Palembang (Ahmad et al., 2007), and East Jakarta (Hardjanti et al., 2015). Therefore, a new strategy of controlling mosquitoes need to develop. It has prompted the researcher to look for environment-friendly, safe for human and animal, target specific for mosquitoes, and cost effective. Another control method such as botanical and microbial insecticide has been applied for last few year (Elumalai et al., 2016). Due to limited use of it in vector control programs, no study reported about resistance.

Plant extract may be alternative source due to it has the abundant bioactive compound. Artemisia vulgaris is the member of the family of Asteraceae. A. vulgaris has height about 1-1.5 meter and release aromatic fragrant. Artemisia distributed throughout the northern temperate regions of Africa, Europe, Asia, and North America. The chemical studies on A. vulgaris indicate that thujone isomer and camphor were determined as the main components in A. Vulgaris from India (Judžentienė & Buzelyte 2006). Compounds like terpenoids and flavonoids are present in this plant. Artemisia extracts also contain artemisinin that has been reported to reduce Plasmodium falciparum gametocyte development, thus reducing transmission of malaria, this fact significant in preventing the spread of resistant strain (Masotti et al., 2012). However, to our knowledge, the mosquito adulticidal activity of A. vulgaris extract has been little investigated.

METHODS

Plant collection

The plant *A. vulgaris* were collected from Tlahap, Kledung, Temanggung, Central Java (7°19'30' S and 110°14' 88' E) in March 2017. Taxonomic identification was issued by Department of Plant Systematic, Biology Faculty, Universitas Gadjah Mada, Yogyakarta, Indonesia.



Figure 1. Artemisia vulgaris from Tlahap, Kledung, Temanggung, Central Java

Mosquitoes rearing

The eggs of *Ae. Aegypti* was collected using about 200 ovitraps in several places in Sleman, Yogyakarta. Eggs were transferred to laboratory Parasitology, Faculty of Veterinary Medicine, Universitas Gadjah Mada. Eggs were hatched using plastic container 20 x 15 x 5 cm3 filled with 800 ml tap water within \pm 24 hours. Larvae were fed with chicken liver and maintained at 28°C, 70-85% relative humidity, with a photoperiod of 14 h light

and ten h dark. Pupae were transferred to a cup (12 cm x diameter 9.8 cm) containing 200 ml clean water covered with a net for adult emergence. The adult was moved into the cage ($20 \times 20 \times 20 \text{ cm}$) and fed using 10% sucrose solution soaked in cotton.

Extraction

A total of 5 kilograms *A. vulgaris* were cleaned and dried using the oven for seven days with temperature 55°C.Total 300 grams dried leaves were grounded into powder by grinding machine. Ethanol 95% (Emsure, Germany) was added to the powder for *A. Vulgaris*maceration. The mixture of *A vulgaris*and ethanol is homogenized for 30 minutes and kept for one day at room temperature. Complete removal of the filtrate was accomplished in vacuum rotary evaporator. The extract then is heated in the water bath at a temperature of 70°C. The final extract was in the pasta form and kept in 4°C until further use.

Adulticidal test using Center for Diseases Control Prevention (CDC) Standard

Weighing one gram of extract dissolved in 100 ml of ethanol for making 1% of the stock solution. To make 10, 50, 100, 500, 1000, 5000, 10000, 50000, and 100000 μ g per bottle. Stock solution diluted using ethanol and transferred to a bottle according to the concentration required. The bottle used has a volume of 250 ml, the bottle was washed using soapy water then rinsed three times and placed in an oven for 20 minutes until dry. One ml of each concentration solution added to the bottle including for control using only ethanol; the bottle rotated gently to make all side coated with the solution, the caps were removed and continuously rolled until the solution completely dry. The bottle was kept from direct sunlight.

Bioassay procedure

The assay was performed using 10-25 F0 mosquitoes (2-5 days old sucrose fed) put into the previous bottle using the aspirator. Every 15 minutes the mosquitoes were observed until two hours long. Mosquitoes were considered died if they could no longer stand, the dead and alive mosquitoes were recorded. This procedure was performed triplicates include the control.

Statistical analysis

The average mortality was subjected to ANOVA using GraphPad Prism 7.00 and probit analysis for calculating lethal dosages causing 50% and 90% mortality (LC50 and LC90) also 95% upper and lower confidence limit.

RESULTS AND DISCUSSION

The adulticidal activity of *A. vulgaris* in the ethanolic extract at various concentration against *Ae. aegypti*was illustrated in figure 2. Observation of present study showed that mortality was increased as the concentration increased in 90, 105 and 120 minutes. For example in 100 mg concentration, mortality

was measured 6,7% and increased to 15% at 500 μ g in 120 minutes. 100% mortality was reached at 10000 μ g. The LC50 and LC90 were determined by using probit analysis (GraphPad Prism 7.0.). The results given in figure 3 showed that LC50 and LC90 were 5790 μ g and 52110 μ g respectively after 120 minutes exposure to the extract. The 95% lower confidence limit is 3331 μ g and upper confidence limit is 9961 μ g. There was no mortality in ethanol control group. *A. vulgaris* significantly (P < 0.05) have adulticidal activity against *Ae. aegypti*.



Figure 2. Adulticidal activity of *A. vulgaris* extract against *Ae. aegypti.*



Figure 3. Graph showed result of probit analysis

Insecticides resistance tends to increase worldwide (Hamid *et al.*, 2017, Ishak *et al.*, 2015, Li *et al.*, 2015, Lima *et al.*, 2011, Saavedra *et al.*, 2007). This problem becomes a dangerous nowadays since arthropod-borne diseases prevention strategy depends on the vector controls. Compounds from natural herb can be used as an alternative insecticide to control the vectors (Govindarajan *et al.*, 2011). Bioinsecticide of plant origin was used against several insect species due to safer to human, without toxic and no residues left in the environment.

The adulticide assay of this study showed activity against adult *Ae. aegypti*. We adapted CDC bottle assay in this experiment to

demonstrate the ability of A. vulgaris in penetrating adult Ae. aegypti and killing the mosquitoes. According to literature survey, lack of published paper about the adulticidal activity of A. vulgaris on Ae. aegypti. However, A. vulgaris have been reported to have larvacidal activity against Ae. Aegypti (Mya et al., 2016, Sundararajan & Kumari, 2017). Another species of Artemisia, A. nilagirica have been reported to have adulticidal activity against Ae. aegypti (Panneerselvam et al., 2012) indicated that A. nilagirica has adulticide activity the LC50 and LC90 values 242.52 and 523.73 ppm. A. annua in the previous report showed that have larvacidal activity, ovicidal and oviposition deterrent (Cheah et al., 2013), A. abrotanum and A. pontica also showed have larvacidal activity (Tabanca et al., 2011). Using Spondia mombin leaves in methanol extract (Ajaegbu et al., 2016) with CDC bioassay procedure showed the LC50 values of 4061.946 µg/bottle in 24 hours post treatment. Ae. aegypti showed restless movement, abnormal wagging, and died inside the bottle contain A. vulgaris extract only within 120 minutes.

Another plant has adulticidal activity using WHO protocol, essential oil *Piper retrofractum Vahl* in acetone showed that LD50 and LD99 of against *Ae. aegypti* (8.86%, 23.21%) and *Culex quinquefasciatus* (6.95% and 17.35%) (Subsuebwong*et al.*, 2016).

Cassiatora leaf extracts against the adulticidal activity of (hexane, chloroform benzene, acetone, and methanol) Ae. aegvptiLC(50) values were 329.82, 307.31, 287.15, 269.57, and 252.03 ppm and LC(90) values were 563.24, 528.33, 496.92, 477.61, and 448.05 ppm (Amerasanet al. 2012). Citrus sinensis orange peel extract against Ae. aegypti LC50 showed value was 320.38ppm (Muruganet al., 2012). Clausena dentata plant extract against Ae. aegypti showed the LC50 and LC90 4.1783 mg/ml and 9.3884 mg/ml (Ramkumar et al., 2015). The obtained results indicate that the extract of A. vulgaris has potential to be developed as an adulticide against Ae. aegypti mosquitoes. However, further studies to evaluate its toxicity need to be conducted. Studies aimed at isolation andidentification of active compounds must be performed. The results of the present study could be used in promoting research aimed at the development of new agents for mosquito

control based on bioactive chemical compounds from indigenous plant sources.

CONCLUSION

The results indicate that the extract of *A. vulgaris* has potential to be developed as an adulticide against *Ae. aegypti* mosquitoes with LC50 and LC90 were 5790 mg and 52110 mg. However, further studies to evaluate its toxicity need to be conducted.

ACKNOWLEDGEMENT

This study was funded by Student Creativity Program and partially funded by INSINAS grant no. 226/SP2H/LT/DRPM/III/2016 by Ministry of Research, Technology, and Higher Education of the Republic of Indonesia.

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